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Global prevalence of *Clostridioides difficile* in 17,148 food samples from 2009 to 2019: a systematic review and meta-analysis

Soroush Borji^{1†}, Sepide Kadivarian^{1†}, Shirin Dashtbin², Sara Kooti³, Ramin Abiri⁴, Hamid Motamedi^{1,7}, Jale Moradi¹, Mosayeb Rostamian^{5*} and Amirhooshang Alvandi^{6*}

Abstract

Background *Clostridioides* (*Clostridium*) *difficile* is an important infectious pathogen, which causes mild-to-severe gastrointestinal infections by creating resistant spores and producing toxins. Spores contaminated foods might be one of the most significant transmission ways of *C. difficile*-associated infections. This systematic review and meta-analysis study were conducted to investigate the prevalence of *C. difficile* in food.

Methods Articles that published the prevalence of *C. difficile* in food in PubMed, Web of Science, and Scopus databases were retrieved using selected keywords between January 2009 and December 2019. Finally, 17,148 food samples from 60 studies from 20 countries were evaluated.

Results The overall prevalence of *C. difficile* in various foods was 6.3%. The highest and lowest levels of *C. difficile* contamination were detected to seafood (10.3%) and side dishes (0.8%), respectively. The prevalence of *C. difficile* was 4% in cooked food, 6.2% in cooked chicken and 10% in cooked seafood.

Conclusions There is still little known concerning the food-borne impact of C. difficile, but the reported contamination might pose a public health risk. Therefore, to improve the food safety and prevent contamination with *C. difficile* spores, it is necessary to observe hygienic issues during foods preparation, cooking and transfer.

Keywords Clostridioides (Clostridium) difficile, Food, Prevalence, Public health

[†]Soroush Borji and Sepide Kadivarian contributed equally to this work

*Correspondence:

Mosayeb Rostamian

- mosayeb.rostamian@gmail.com
- Amirhooshang Alvandi ah_alvandi@kums.ac.ir
- ¹ Department of Microbiology, School of Medicine, Kermanshah
- University of Medical Sciences, Kermanshah, Iran
- ² Department of Microbiology, School of Medicine, Iran University of Medical Sciences, Tehran, Iran
- ³ Behbahan Faculty of Medical Sciences, Behbahan, Iran

⁴ Fertility and Infertility Research Center, Research Institute for Health Technology, Kermanshah University of Medical Sciences, Kermanshah, Iran

⁵ Infectious Diseases Research Center, Health Institute, Kermanshah University of Medical Sciences, Kermanshah Postal Code: 6714415333, Iran

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⁶ Department of Microbiology, School of Medicine, Medical Technology Research Center, Research Institute for Health Technology, Kermanshah University of Medical Sciences, Kermanshah Postal Code: 6714415333, Iran

⁷ Student Research Committee, School of Medicine, Kermanshah University of Medical Sciences, Kermanshah, Iran

Background

In the mid-1970s, the gram-positive and anaerobic bacterium Clostridioides difficile (formerly known as Clostridium difficile) was found as a common cause for nosocomial infection and a major cause of antibioticassociated diarrhea [1-3]. By forming resistant spores and the ability of producing toxins, C. difficile is responsible for a diverse group of infection, from mild and self-limiting gastrointestinal infections to severe life threatening infections, like toxic megacolon [4, 5]. C. difficile infection (CDI) is associated with significant mortality and increased healthcare costs in the world [6-9]. C. difficile is basically a nosocomial pathogen, but the prevalence of community-acquired CDI seems to be increasing [10, 11]. Prevalence of C. difficile contamination in food is high, and a wide range of foods are contaminated by C. difficile [12, 13]. Therefore, consumption of C. difficile contaminated food is a risk factor for transmission of this infection in community, and one of the most important route of transmitting could be contaminated food by C. difficile spore [14, 15]. The presence of C. difficile in sewage-treatment plants might be a major reason of its community acquisition, transmission to food, and ultimately food contamination [14, 16]. This issue demands more attention to this health-threatening pathogen.

The main aims of this systematic and meta-analysis study were (i) to investigate the prevalence of *C. difficile* in different types of food and compare them with each other, (ii) to determine the frequency of toxin genes, (iii) to assay the relationship of toxin genes with the prevalence of *C. difficile*, and (iv) to evaluate the phenotypic and genotypic diagnostic methods from 17,148 food samples.

Methods

Literature search

Published studies from January 2009 to December 2019 were retrieved from four main databases including Web of sciences, Scopus, PubMed, and Google Scholar by applying the following keywords: "clostridia", "*Clostridium spp*", "*Clostridium difficile*", "*Clostridioides difficile*", "*C. difficile*", "antibiotic resistance", "food contamination", "toxinotype", "ribotype", and "toxin genes" alone or combined with "AND" and/or "OR" operators. To conduct the present study, Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA) guideline were considered [17].

Inclusion/exclusion criteria

All cross-sectional studies focusing on the prevalence of *C. difficile* contamination in food samples were included. Short communications, cohort studies, clinical trials,

letter to editors, narrative or systematic reviews, and the non-English articles were excluded.

Selection of studies and data gathering

The text of all included studies was accurately read by two independent authors, and in case of any discrepancy, the issue was discussed by other authors to be resolved. The following characteristics of each study were collected: first author, year of publication, sampling year, the location of the study, detection methods, sample type, sample size, the number of detected *C. difficile*, toxinotypes, ribotypes, toxin genes, antibiotics used, number of resistance isolates, and the method of antibiotic susceptibility assay.

Data analysis

Data analyses were performed using Comprehensive Meta-Analysis software, V2.2.064. The C. difficile prevalence in different food samples and the prevalence of toxinotype and toxin genes, and antibiotic resistance rate in the C. difficile isolates were shown with event rate and a 95% confidence interval (CI). The random-effects model was chosen for meta-analyses, and several subgroup analyses were conducted to evaluate the source of heterogeneity based on the continent, country, sample types and the sampling periods of time. Using a random-effects model, risk ratios for each sample type were calculated to quantify the differences and rank the sample types based on the risk. The Q test and I2 statistic were applied to measure any possible heterogeneity between the studies. The publication bias was evaluated by conducting Egger weighted regression test. In all analyses, the significate threshold was < 0.05 (p value < 0.05).

Results

Search results

In total, 2202 studies were recovered after accurate searching in the databases using the aforementioned key words. Among them, 1026 papers were non-duplicated articles and were considered in the study. After title/ abstract screening, 116 studies remained. For eligibility, 79 studies were assessed by full-text reading. Sixty studies remained for final qualitative and meta-analysis. The diagram of our search strategy is given in Fig. 1, and the extracted characteristics of the studies are shown in Table 1.

The pooled prevalence of C. difficile in food samples

To analyze the pooled prevalence of *C. difficile* in food samples, 60 studies were used in a random-effects model. The event rate, which was the number of *C. difficile* cases over the number of samples, was applied as the effect size index. The overall pooled prevalence of

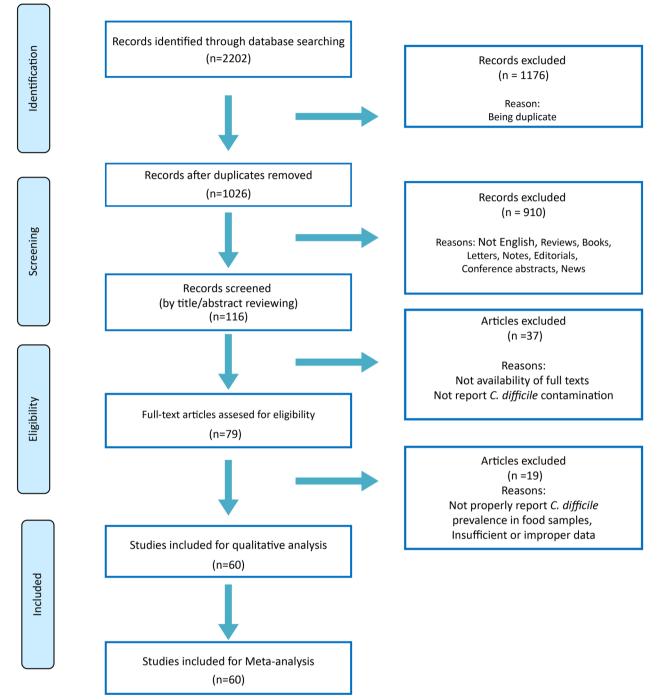


Fig. 1 PRISMA flow diagram of study selection

C. difficile in food samples was estimated to be 6.3% (CI 95%: 4.8–8.2) (Fig. 2). The lowest and highest *C. difficile* prevalence was observed in Shaughnessy et al. and Romano et al. reports with 0.1% and 66.7% prevalence, respectively (Fig. 2). The *Q*-value was 1049.1

which was much higher than the number of studies minus 1 (60-1=59), that reject the null hypothesis and showed a significant heterogeneity between studies. The I² statistics indicated that 94.4% of the variances reflect true variances between studies.

Table 1 The characteristics of the studies

Study and Reference	Published year	Sampling year	Country	Continent	Detection method	Number of samples	Confirmed C. <i>difficile</i> No	Sample type	Sample size of each category	C.d isolates of each category	Antibiotic susceptibility assay	Antibiotic resistance*	Toxin genes*	Toxinotype*	Ribotype*
Abdel-Glil et al. [41]	2018	2014–2015	Egypt	Africa	PCR	150	0	R-meat	150	0	I	I	I	I	I
Kouassi et al. [42]	2014	2009–2010	Cote d'Ivoire		PCR	395	49	C-meat/ Ham	395	49	DD	+	I	I	I
Esfandiari	2014	2012	Iran	Asia	PCR	211	6	R-meat	166	6	Ι	I	+	I	+
I1 [43]								Soy	14	0					
								S-dishes	17	0					
								Veg	14	0					
Esfandiari et al2 [44]	I	2014	Iran		PCR	200	œ	R-meat	200	œ	I	I	+	I	+
Esfandiari-	2013	2012-2013	Iran		I	211	22	R-meat	110	14	I	Ι	I	I	Ι
l3 [45]								Veg	70	8					
								Soy	14	0					
								S-dishes	17	0					
Esfandiari et al4 [46]	2014	I	Iran		PCR	100	12	R-meat	100	12	I	I	+	I	I
Hasanzade et al1 [47]	2013	I	Iran		I	240	25	R-poultry	240	25	I	I	I	I	I
Hasanzadeh et al2 [48]	2013	I	Iran		I	120	19	R-poultry	120	19	I	I	I	I	I
Kherad- mand et al. [49]	2017	2014–2015	Iran		PCR	100	30	R-meat	100	30	I	I	+	I	I
	2016	2013	Australia		PCR	300	76	R-meat	300	76	I	I	I	I	I
Kochak- khani et al. [51]	2017	2015	Iran		RT-PCR	60	œ	Salad	60	œ	DD	+	I	I	I
Lee et al. [52]	2018	2013-2014	South Korea		I	415	45	R-meat R-poultry	266 149	20 25	E-test	+	+	I	I
Lim et al. [53]	2018	2015	Australia		PCR	300	30	Veg	300	30	I	I	+	I	+
Nayebpour et al.[54]	2018	2017	Iran		PCR	820	26	Seafood	820	26	DD	+	+	I	I
Rahimi et al1 [55]	2014	2012	Iran		PCR	660	13	R-meat	660	13	DD	+	+	I	+
Rahimi	2015	2013	Iran		PCR	368	5	Salad	248	4	DD	+	+	I	I
[oc] 7-:1								Milk/Dairy	50	-					
								S-dishes	70	0					

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Table 1	Table 1 (continued)														
Study and Reference	Published year	Sampling year	Country	Continent	Detection method	Number of samples	Confirmed C. <i>difficile</i> No	Sample type	Sample size of each category	C.d isolates of each category	Antibiotic susceptibility assay	Antibiotic resistance*	Toxin genes*	Toxinotype*	Ribotype*
Rahimi et al3 [57]	2015	2012	Iran		PCR	570	9	C-meat/ Ham	100	-	I	I	+	1	1
								R-meat	105	5					
								R-poultry	150	0					
								RTE meat	170	0					
Razmyar et al. [58]	2017	2014	Iran		PCR	65	10	R-poultry	65	10	I	I	+	I	I
Rivas et al.	2019	2013-2014	New		RT-PCR	170	-	RTE meat	55	-	E-test	+	+	I	+
[29]			Zealand					Pet Food	9	0					
								Veg	9	0					
								R-meat	55	0					
								R-poultry	16	0					
								Milk/Dairy	2	0					
								Salad	27	0					
								Seafood	c.	0					
Wu et al. [60]	2017	2015	Taiwan		PCR	302	67	R-meat	302	67	E-test	I	+	I	+
Yamoudy et al. [61]	2015	2013	Iran		PCR	106	9	Salad	106	9	I	I	+	I	I
Zamani et al. [62]	2019	2013-2014	Iran		PCR	30	2	R-meat	30	2	I	I	I	I	I
Curry et al. [63]	2012	2011–2012	USA	Central/ North America	I	102	13	R-meat	102	13	I	I	+	I	+
Gomez et al. [64]	2013	2009–2010	Costa Rica		PCR	200	4	R-meat R-poultry	133 67	e 1	E-test	+	+	I	+
Han et al. [65]	2018	2014-2015	NSA		RT-PCR	297	41	Veg	297	41	MIC	+	+	I	+
Harvey et al1 [66]	2011	2004–2009	USA		PCR	243	20	R-meat	221	20	E-test	I	+	+	I
Harvey et al2 [67]	2011	2010	NSA		PCR	32	7	R-poultry	32	7	E-test	+	+	+	I
Hawken et al.1 [68]	2013	I	Canada		PCR	80	œ	R-meat	80	œ	I	I	+	I	+
Hawken et al2 [69]	2013	I	Canada		PCR	78	4	R-meat	78	4	I	I	+	+	+
Houser et al.	2010	I	USA		RT-PCR, PCR	80	0	R-meat	36	0	I	I	I	I	I
[0/]								R-poultry	17	0					
								Milk/Dairy	27	0					

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Study and Reference	Published year	Sampling year	Country	Continent	Detection method	Number of samples	Confirmed C. <i>difficile</i> No	Sample type	Sample size of each category	C.d isolates of each category	Antibiotic susceptibility assay	Antibiotic resistance*	Toxin genes*	Toxinotype*	Ribotype*
Kalchay- anand et al. [71]	2013	2006-2007	USA		PCR	2427	18	R-meat	2427	18	MIC	+	+	+	+
Kwon et al. [72]	2016	2011-2012	USA		PCR	910	7	S-dishes Veg R-meat R-poultry Milk/Dainv	380 831 308 142 210		I	I	+	I	+
Metcalf et al1 [73]	2010	2007-2008	Canada		PCR	393	7	R-meat C-meat/ Ham	2 - 	o 4 m	E-test	+	+	+	+
Metcalf et al2 [74]	2011	2010	Canada		PCR	119	5	Seafood C-seafood	28 10	4 ←	E-test	+	+	+	+
Mooyottu et al. [75]	2015	I	USA		PCR	300	2	R-meat R-poultry	200 100	2	E-test	+	+	I	I
Norman et al. [76]	2014	2012	USA		PCR	67	c,	Seafood	67	m	E-test	+	+	+	I
Shaugh- nessy et al. [77]	2018	2011-2012	USA		PCR	342	0	R-meat R-poultry	256 109	0 0	I	I	I	I	I
Songer et al. [28]	2009	2007	USA		PCR	88	37	R-meat RTE meat	65 23	26 11	E-test	+	+	+	+
Varshney et al. [78]	2014	2011-2012	NSA		RT-PCR, PCR	303	31	R-meat R-poultry	150 153	14	E-test	+	+	I	+
Visser et al. [79]	2012	2007	Canada		PCR	48	ć	R-meat	48	m	E-test	+	+	Ι	I
Weese et al1 [80]	2009	2008	Canada		PCR	230	28	R-meat	230	28	I	I	+	+	+
Weese et al2 [81]	2010	2008–2009	Canada		PCR	203	26	R-poultry	203	26	I	I	+	I	+
Agnoletti et al. [<mark>82</mark>]	2019	2015-2017	Italy	Europe	RT-PCR, PCR	702	113	Seafood	702	113	E-test	+	+	I	I
Abercron et al. [31]	2009	2008	Sweden		I	82	7	R-meat C-poultry C-meat/ Ham	65 4 13	0 0 7	I	I	+	I	I
Bakri et al. 1831	2009	2008	UK		PCR	40	e	Veg	48	2	E-test	+	+	I	+

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study and rublished sampleference year year de Boer 2011 2008. Et al. [27] 2010 [84] Et al. 2013 2010 [84] [84] 2018 2013 [29]	-2014 -2014	~	Сонциент	method	number of samples	Confirmed C. <i>difficile</i>	type	sampie size of each	isolates	Antipiotic susceptibility	Antipiotic resistance*	genes*	loxinotype	kipotype
oer 2011 .[27] .rt et al. 2013 z et al. 2018		et alt an				No		category	or eacn category	assay				
rt et al. 2013 z et al. 2018		Nether- land		RT-PCR, PCR	500	œ	R-meat R-noultry	243 257	1 2	I	I	+	1	+
z et al. 2018		France		PCR	104	m	Salad	, c 7 60	, 7	QQ	+	I	I	+
z et al. 2018							Veg	44	-					
		Turkey		I	101	2	C-meat/ Ham	25	2	E-test	+	I	I	I
							R-meat	31	0					
							R-poultry	27	0					
							C-poultry	12	0					
							RTE meat	9	0					
Guran et al. 2015 2012 [85]	2012–2013 T	Turkey		PCR	310	25	R-poultry	310	25	I	I	+	I	I
Hampikyan 2018 – et al. [86]	F	Turkey		PCR	555	161	R-meat	555	161	E-test	+	+	I	+
tl et al. 2010	2007-2008	Austria		PCR	120	c	R-meat	70	č	E-test	+	+	I	+
[87]							Milk/Dairy	50	0					
Pasquale 2012 2010 et al. [36]	2010-2011	Italy		PCR	52	26	Seafood	52	26	I	I	+	+	+
Primavilla 2019 2016 et al. [88]	2016-2017	Italy		RT-PCR	350	2	C-meat/ Ham	130	-	E-test	+	+	+	+
							Veg	54	-					
							S-dishes	166	0					
Rodriguez 2014 2012 et al1 [89]		Belgium		PCR	240	œ	R-meat	240	∞	E-test, DD	+	+	I	+
Rodriguez 2015 2013 et al2 [90]		Belgium		PCR	188	-	I	I	-	I	I	+	I	+
Rodriguez 2013 2011 et al3 [24]	2011-2012 E	Belgium		PCR	201	15	R-meat	201	15	I	I	+	I	+
Romano 2018 2009 et al. [91]	2009–2013	Italy		PCR	6	9	Milk/Dairy Salad	3 Q	3 Q	I	I	+	I	+
Troian et al. 2015 2012 [35]	2012-2014	Italy		PCR	925	36	Seafood	925	36	E-test	+	+	I	+
Valerija et al. 2018 2014 [20]	2014-2017 5	Slovenia		PCR	154	28	Veg	154	28	I	I	I	+	+
Pires et al. 2018 2017 [92]		Brazil S	South America	RT-PCR	80	0	R-meat	80	0	I	I	I	I	I

Study name		Stat	istics for each	n study			Event	rate and 95% C	l.	
	Event rate	Lower limit	Upper limit	Z-Value	p-Value					
Primavilla et al.	0.006	0.001	0.023	7.275-	0.000	1		+	1	1
Agnoletti et al.	0.161	0.136	0.190	16.076-	0.000					
Abdel-Glil et al.	0.003	0.000	0.051	4.029-	0.000			÷		
Pires et al.	0.006	0.000	0.091	3.582-	0.000			+		
Songer et al.	0.420	0.322	0.526	1.486-	0.137			-		
Bakri et al.	0.075	0.024	0.208	4.185-	0.000			-		
Jobstl et al.	0.025	0.008	0.075	6.266-	0.000			•		
Rivas et al.	0.006	0.001	0.041	5.115-	0.000			ŧ		
Gomez et al.	0.020	0.008	0.052	7.705-	0.000			•		
Kalchayanand et al.	0.007	0.005	0.012	20.697-	0.000			•		
Abercron et al.	0.024	0.006	0.092	5.153-	0.000			+		
Eckert et al.	0.029	0.009	0.086	6.002-	0.000			•		
Visser et al.	0.063	0.020	0.177	4.542-	0.000			•		
Harvey et al1	0.082	0.054	0.124	10.331-	0.000					
Weese et al1	0.122	0.085	0.171	9.799-	0.000					
Curry et al.	0.127	0.075	0.207	6.479-	0.000			_ ⊢	1	
Metcalf et al1	0.018	0.009	0.037	10.514-	0.000			Ī	1	
de Boer et al.	0.016	0.008	0.032	11.557-	0.000			Ē		
Esfandiari et al1	0.043	0.022	0.080	9.132-	0.000					
Hawken et al.1	0.100	0.051	0.187	5.896-	0.000					
Hawken et al2	0.051	0.019	0.129	5.684-	0.000			_ ⊺ _		
Harvey et al2	0.219	0.108	0.393	2.977-	0.003					
Razmyar et al.	0.154	0.085	0.263	4.959- 8.807-	0.000					
Esfandiari et al2 Ersoz et al.	0.040 0.020	0.020 0.005	0.078 0.076	5.463-	0.000 0.000			1		
Kouassi et al.	0.020	0.005	0.160	12.806-	0.000			T_		
Kwon et al.	0.002	0.000	0.009	8.643-	0.000			1-		
Lee et al.	0.002	0.082	0.003	13.345-	0.000			T_		
Lim et al.	0.100	0.071	0.139	11.417-	0.000					
Metcalf et al2	0.042	0.018	0.097	6.843-	0.000					
Mooyottu et al.	0.007	0.002	0.026	7.053-	0.000			ſ		
Norman et al.	0.045	0.015	0.130	5.181-	0.000			L.		
Pasquale et al.	0.500	0.367	0.633	0.000	1.000			_ _		
Rahimi et al1	0.020	0.011	0.034	13.949-	0.000					
Rodriguez et al1	0.033	0.017	0.065	9.364-	0.000			T I		
Rodriguez et al2	0.005	0.001	0.037	5.217-	0.000			÷.		
Romano et al.	0.667	0.333	0.889	0.980	0.327			−		
Shaughnessy et al.	0.001	0.000	0.023	4.614-	0.000			+		
Valerija et al.	0.182	0.129	0.251	7.199-	0.000					
Troian et al.	0.039	0.028	0.053	18.861-	0.000			•		
Varshney et al.	0.102	0.073	0.142	11.457-	0.000			-		
Weese et al2	0.128	0.089	0.181	9.132-	0.000			-		
Wu et al.	0.222	0.179	0.272	9.061-	0.000			■		
Zamani et al.	0.067	0.017	0.231	3.606-	0.000			-		
Rahimi et al2	0.014	0.006	0.032	9.516-	0.000			+	1	
Esfandiari-3	0.104	0.070	0.153	9.547-	0.000			■		
Yamoudy et al.	0.057	0.026	0.120	6.694-	0.000			•		
Esfandiari et al4	0.120	0.069	0.200	6.475-	0.000			-	1	
Kheradmand et al.	0.300	0.218	0.397	3.883-	0.000			-		
Guran et al.	0.081	0.055	0.117	11.667-	0.000			-		
Hampikyan et al.	0.290	0.254	0.329	9.568-	0.000			•	1	
Han et al.	0.138	0.103	0.182	10.888-	0.000			-	1	
Houser et al.	0.006	0.000	0.091	3.582-	0.000			t t	1	
Rahimi et al3	0.011	0.005	0.023	11.070-	0.000			•	1	
Hasanzade et al1	0.104	0.071	0.150	10.183-	0.000					
Hasanzadeh et al2	0.158	0.103	0.235	6.681-	0.000				1	
Kochakkhani et al.	0.133	0.068	0.245	4.929-	0.000			_=-		
Nayebpour et al.	0.032	0.022	0.046	17.155-	0.000			•	1	
Knight et al.	0.253	0.207	0.306	8.143-	0.000			L■		
Rodriguez et al3	0.075	0.045	0.120	9.380-	0.000				1	
	0.063	0.048	0.082	18.431-	0.000	I	I	l+	I	
						-2.00	-1.00	0.00 1	.00	2.00
							Favours A	Four	ours B	
							Favours A	Favo		

C. difficile pooled prevalence

Q: 1049.1 I-squared: 94.4

Fig. 2 *C. difficile* pooled prevalence. The overall pooled prevalence of *C. difficile* in food samples was estimated to be 6.3% (Cl 95%: 4.8–8.2). The lowest and highest *C. difficile* prevalence was observed in Shaughnessy et al. and Romano et al. reports with 0.1% and 66.7% prevalence, respectively

Subgroup	Number of studies	Prevalence (%)	Lower limit	Upper limit	Z-value	<i>p</i> value	l ²
Based on the continent							
Africa	2	4.8	0.8	24	- 3.2	0.001	85.6
Asia	20	7.4	4.6	11.7	- 9.9	0.000	93.8
Central/North America	20	5.1	3.1	8.4	- 10.9	0.000	94.1
Europe	17	6.8	3.9	11.3	- 9.1	0.000	95.6
South America	1	0.6	0.08	17.0	- 2.8	0.004	0.0
Overall	60	6.2	4.7	8.2	- 17.8	0.000	94.4
Test of heterogeneity between	subgroups: Q-valu	ie:3.095, <i>p</i> value: 0.542					
Based on the sampling year							
TF1 (2004-the end of 2008)	8	5.1	2.3	10.9	- 6.9	0.000	97.0
TF2 (2009-the end of 2013)	23	5.7	3.6	9	- 11.2	0.000	93.6
TF3 (2014 <u>≤</u>)	13	8.4	4.6	15.1	- 7.1	0.000	92.4
Overall	44	6.3	4.5	8.7	- 14.9	0.000	94.4
Test of heterogeneity between	n subgroups: Q-valu	ie: 1.372, <i>p</i> value: 0.504					
Based on the sample types							
C-meat/Ham	5	4.0	1.3	12.1	- 5.2	0.000	74.2
C-poultry	2	6.2	0.5	44.8	- 2.1	0.034	0.0
C-seafood	1	10.0	0.6	67.6	— 1.5	0.144	0.0
Milk/Dairy	6	4.5	1.2	15.7	- 4.3	0.000	77.6
Pet Food	1	7.1	0.2	72.8	- 1.4	0.157	0.0
R-meat	35	5.6	4.0	8.4	— 13.6	0.000	94.3
R-poultry	17	6.1	3.4	10.7	- 8.7	0.000	74.5
RTE meat	4	7.9	2.0	26.5	- 3.3	0.001	88.6
Salad	7	6.1	2.4	14.9	- 5.4	0.000	75.0
S-dishes	5	0.8	0.2	3.3	- 6.4	0.000	0.0
Seafood	7	10.3	4.6	21.4	- 4.9	0.000	96.4
Soy	2	3.3	0.3	29.1	- 2.7	0.008	0.0
Veg	10	5.7	2.6	11.8	- 6.9	0.000	77.7
Overall	102	5.7	4.5	7.3	- 21.6	0.000	90.2
Test of heterogeneity between	n subgroups: Q-valu	ie: 10.657, <i>p</i> value: 0.55	7				

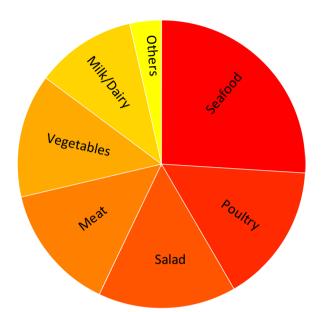
Table 2 Subgroup analysis of C. difficile prevalence based on the studies continent, the sampling year and the sample types

Subgroup analysis of *C. difficile* prevalence based on the study continent, the year of sampling and the sample types

To subgroup analysis of *C. difficile* prevalence in food samples was performed based on the study continent, in which the 60 studies were divided into the following subgroups: Africa (two studies), Asia (20 studies), Central/North America (20 studies), Europe (17 studies), and South America (one study). Difference in prevalence of *C. difficile* isolated from food samples in different continents was not significant (Table 2).

To subgroup analysis of *C. difficile* food prevalence based on the sampling year, three time frames were used as follows: TF1 (2004-the end of 2008), TF2 (2009the end of 2013), and TF3 (2014 \leq). Considering these time frames, 44 studies were used for a random-effects model subgroup analysis. No statistically significant difference was observed between time frame subgroups (Table 2).

To subgroup analysis of *C. difficile* food prevalence based on the sample type, the following subgroups were used: raw meat (R-meat), cooked meat/Hamburger (C-meat/Ham), poultry raw meat (R-poultry), cooked poultry (C-poultry), raw seafood/fish (Seafood), cooked seafood/fish (C-seafood), vegetables (Veg.), ready-toeat meat (RTE meat), Milk/Dairy, salad, soy, side dishes (S-dishes), and pet food. The prevalence of *C. difficile* in each sample type is presented in Table 2. The highest and lowest prevalence were 10.3% and 0.8%, which were seen in Seafood and S-dishes sample types, respectively (Table 2). Although there were some differences in *C. difficile* prevalence of different sample types, no significant heterogeneity was observed between groups (*Q*-value: 10.657, *p* value: 0.557) (Table 2).



Sample type	Number of reports	C. difficile Prevalence (%)
Seafood	8	10.3
Poultry	19	6.2
Salad	7	6.1
Meat	44	5.6
Vegetables	10	5.5
Milk/Dairy	6	4.5
Others	8	1.4

Fig. 3 The prevalence of *C. difficile* in different sample types

For better presentation of the results, in another arrangement, the studies were divided to more general groups based on sample types as follows: meat, poultry, seafood, vegetables, salad, milk/diary, and others (S-dishes, soy, pet food) (Fig. 3).

For presenting each sample type in each country, more subgroup analyses were performed. The summary results of these analyses are shown in Fig. 4. Also, risk ratios were obtained using the extracted data. Based on the ranking of the risk ratio, S-dishes as a reference and was the lowest source of *C. difficile* and seafood, RTE meat, C-poultry, salad, R-poultry and R-meat had highest risks. Compared to S-dishes, the probability of contamination of seafood with CD was 12.88 times higher than S-dishes, and the risk of contamination of RTE meat, C-poultry, salad, R-poultry and R-meat obtained 9.75, 7.75, 7.63, 7.63 and 7.0 times more than S-dishes, respectively (Fig. 5).

Prevalence of *C. difficile* ribotype, toxinotypes and toxin genes

According to a very diverse reported ribotypes, it was impossible to analyze the pooled prevalence of the ribotypes; this parameter is represented in Additional file 1: Table S1 without further analysis.

The most frequent toxinotypes of *C. difficile* were toxinotype 0, III, and V. As it is shown in Table 3, the toxinotype V was more prevalent comparing to other two toxinotypes, and there was a significant heterogeneity between the toxinotypes (*Q*-value: 9.725, *p* value: 0.008) (Table 3).

The toxin genes that were reported in more than one study include genes A, B, CTD, tcdC, tcdC18, tcdC39, tcdC117, and cdtA. The toxin genes of A and B were the most frequent, and genes tcdC18 and tcdC117 were the lowest frequent genes studied (Table 3). There was also significant heterogeneity between the studied genes (*Q*-value: 58.9, *p* value: 0.000) (Table 3).

As shown in Table 4, toxin type 0 in which pathogenic strains were located shows a higher prevalence in seafood samples. While the prevalence of toxin types 3 and 5 was higher in RTE meat and R-poultry. As shown in Table 5, the highest prevalence of toxin genes A, B, and CDT was observed in RTE meat samples. Compared to other samples, Milk/Dairy and Salad rank after RTE meat in terms of the high prevalence of genes A and B toxins.'

Publication bias

The publication bias was checked based on the pooled prevalence of *C. difficile* isolates in food samples. The Egger's linear regression test result showed a significant publication bias in the included studies (p value < 0.0001).

Discussion

Consuming the contaminated raw and cooked foods with C. difficile spore might be an important route of its transmission [18-20]. Food contamination has played an important role in epidemiology of some infectious diseases, but little information is available about the global frequency of C. difficile in food products [21, 22]. The present study analyzed the distribution of C. difficile in 60 studies published from 2009 to 2019 in 17,148 food samples. The results showed that the overall prevalence of *C*. *difficile* in all food samples was 6.3%, with the lowest and highest prevalence of C. difficile were 0.1% and 66.7%, respectively. In a systematic review study, Rodriguez-Palacios and colleagues reported the 4.1% prevalence of C. difficile in human diet samples during 1981 to 2018 [21]. Comparing to the results presented in this study, it seems that the reported prevalence of the bacterium

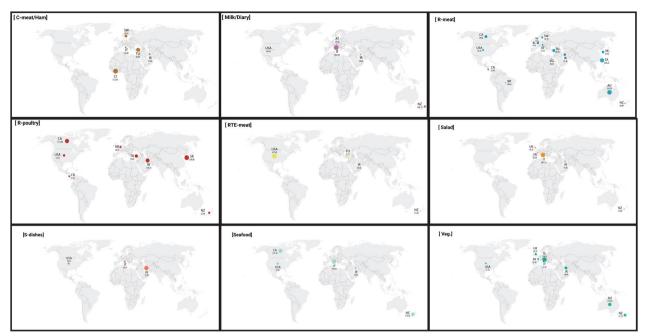


Fig. 4 The prevalence of C. difficile in different sample types in each country. Each sample type is shown in a separate box. The overall prevalence of C. difficile in each country is presented with circles, and the real numbers of prevalence (in percentage) are also presented in parenthesis. EG Egypt, CI Cote d'Ivoire, AT Austria, IR Iran, SK South Korea, TA Taiwan, NZ New Zealand, CA Canada, CR Costa Rica, USA United States of America, AT Austria, BL Belgium, FR France, IT Italy, NE Netherland, Slovenia, SW Sweden, TU Turkey, UK United Kingdom, BR Brazil

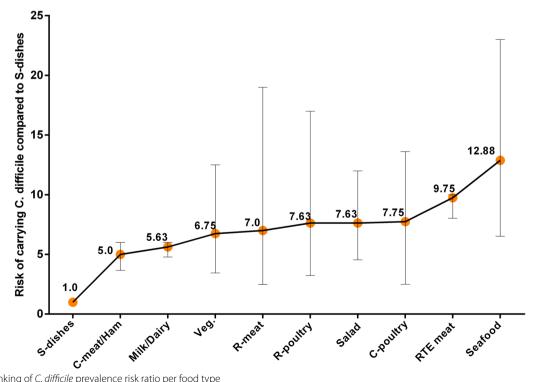


Fig. 5 Ranking of C. difficile prevalence risk ratio per food type

Toxinotype	Number of studies	Prevalence (%)	Lower limit	Upper limit	Z-value	<i>p</i> value	l ²
<i>C. difficile</i> toxinoty	/pes						
0	4	15.6	4.5	42.2	- 2.4	0.016	71.9
111	5	22.7	8.3	48.8	- 2.0	0.041	58.4
V	9	64.5	41.9	82.1	1.3	0.205	79.2
Overall	18	39.1	25.2	54.9	- 1.4	0.175	79.9
Test of heterogen	eity between subgro	oups: Q value: 9.725, p val	ue: 0.008				
Toxin genes	Number of studies	Prevalence (%)	Lower limit	Upper limit	Z-value	p value	l ²
C. difficile toxin ge	enes detected by mo	lecular methods					
А	37	76.8	68.1	83.8	5.3	0.000	64.6
В	37	75.9	66.7	83.2	5.0	0.000	68.7
cdtA	3	28.8	10.1	59.2	- 1.4	0.165	0.0
CTD	27	49.6	36.9	62.3	-0.1	0.953	79.0
tcdC	6	41.7	20.6	66.3	- 0.6	0.516	72.6
tcdC117	4	17.6	5.3	44.7	- 2.3	0.023	63.5
tcdC18	9	19.5	9.6	35.5	- 3.4	0.001	0.0
tcdC39	9	67.4	48.9	81.7	1.9	0.064	68.7
Overall	132	61.7	56.2	67.0	4.1	0.000	74.7
Test of heterogen	eity between subgro	oups: Q-value: 58.9, p valu	e: 0.000				

Table 3 The prevalence of C. difficile toxinotypes and toxin genes

Table 4 The prevalence of C. difficile toxinotypes in each sample type

Toxinotype	Sample type	Number of studies	Prevalence (%)	Lower limit	Upper limit	Z-value	<i>p</i> value	ľ
0	R-meat	3	8.81	3.32	21.35	- 4.43	0.00	0.000
	Seafood	1	42.31	25.20	61.49	- 0.78	0.43	0.000
	Overall	4	26.05	15.91	39.60	- 3.291	0.001	71.943
Test of heteroge	eneity between subg	groups: Q-value: 9	.433, p value: 0.002					
111	R-meat	4	19.93	7.55	43.11	- 2.45	0.01	64.128
	RTE meat	1	36.36	14.33	66.12	- 0.89	0.37	0.000
	Overall	5	26.58	13.69	45.24	- 2.413	0.016	58.436
Test of heteroge	eneity between subg	groups: <i>Q</i> -value: 0	.966, <i>p</i> value: 0.326					
V	R-meat	5	75.65	49.87	90.66	1.95	0.05	75.058
	R-poultry	1	93.75	46.14	99.62	1.85	0.06	0.000
	RTE meat	1	63.64	33.88	85.67	0.89	0.37	0.000
	Seafood	2	38.03	3.87	90.33	- 0.35	0.73	79.274
	Overall	9	70.95	53.09	84.05	2.275	0.023	78.361
Test of heteroge	eneity between subg	groups: Q-value: 2	.99, <i>p</i> value: 0.393					

in these two studies is quiet the same. Taken together, the overall *C. difficile* prevalence in food samples in the world seems to be less than 10%, but it is relatively high and should not be undermined.

Significant heterogeneity was observed between the studies that indicated different prevalence of *C. difficile* in different parts of the world. However, in addition to real differences in *C. difficile* prevalence, the observed

heterogeneity may be due to different seasons of sampling, temperatures and geographical conditions, the quality of studies, the sensitivity of detection methods, etc. [23]. Although the frequency of *C. difficile* varied in food samples from different continents, these differences were not statistically significant. The prevalence of *C. difficile* in Asia and Europe was almost the same, but it was lower in Africa and North/Central America

Toxin gene	Sample type	Number of studies	Prevalence (%)	Lower limit	Upper limit	Z-value	<i>p</i> value	l ²
A	Milk/Dairy	1	92.86	42.28	99.57	1.75	0.08	0.00
	R-meat	21	82.84	70.03	90.89	4.25	0.00	72.93
	R-poultry	4	66.43	44.94	82.75	1.51	0.13	28.75
	RTE meat	1	95.83	57.54	99.74	2.17	0.03	0.00
	Salad	2	87.50	46.27	98.27	1.82	0.07	0.00
	Seafood	6	59.70	50.15	68.56	1.99	0.05	28.90
	Veg	2	79.71	57.76	91.86	2.54	0.01	13.03
	Overall	37	68.76	62.02	74.79	5.180	0.000	64.57
Test of heterogene	eity between subgrou	ups: Q-value: 15.0	1, <i>p</i> value: 0.020					
В	Milk/Dairy	1	92.86	42.28	99.57	1.75	0.08	0.00
	R-meat	21	81.53	66.35	90.80	3.61	0.00	75.38
	R-poultry	4	61.19	40.70	78.36	1.07	0.28	26.09
	RTE meat	1	95.83	57.54	99.74	2.17	0.03	0.00
	Salad	2	87.50	46.27	98.27	1.82	0.07	0.00
	Seafood	6	56.78	43.42	69.22	0.99	0.32	57.52
	Veg	2	95.61	58.20	99.71	2.20	0.03	43.95
	Overall	37	68.38	59.75	75.91	4.016	0.000	68.68
Test of heterogene	eity between subgrou							
CDT	Milk/Dairy	1	16.67	2.28	63.13	- 1.47	0.14	0.00
	R-meat	14	58.10	35.39	77.83	0.69	0.49	70.99
	R-poultry	4	49.23	11.15	88.22	- 0.03	0.98	78.35
	RTE meat	1	95.83	57.54	99.74	2.17	0.03	0.00
	Salad	2	87.50	46.27	98.27	1.82	0.03	0.00
	Seafood	5	28.58	11.54	55.12	- 1.60	0.07	79.94
	Overall	27	51.16	36.47	65.66	0.152	0.879	79.02
Tost of botorogon	eity between subgrou			50.47	05.00	0.152	0.079	7 9.02
tcdC18	R-meat	7	17.29	10.83	26.45	- 5.65	0.00	0.00
	R-poultry	1	23.53	9.12	48.55	- 2.06	0.00	0.00
	RTE meat	1	36.36	14.33	46.33 66.12	- 2.00 - 0.89	0.04	0.00
	Overall	9	20.35	13.96	28.68		0.000	0.00
Tast of botorogon	eity between subgrou	-		13.90	20.00	- 5.893	0.000	0.00
tcdC39		1ps: Q-value: 2.28, 6	63.98	39.88	82.63	1 1 4	0.25	78.03
	R-meat					1.14		
	R-poultry	1	76.47	51.45	90.88	2.06	0.04	0.00
	RTE meat	1	63.64	33.88	85.67	0.89	0.37	0.00
	Seafood	1	91.67	37.82	99.50	1.62	0.10	0.00
	Overall	9	69.81	55.46	81.12	2.654	0.008	68.67
-	eity between subgrou							
tcdC	R-meat	4	42.31	9.65	83.43	- 0.32	0.75	74.84
	R-poultry	1	70.00	37.63	90.02	1.23	0.22	0.00
	Seafood	1	23.08	10.75	42.76	- 2.59	0.01	0.00
	Overall	6	37.08	22.57	54.37	- 1.472	0.141	72.61
	eity between subgrou	ıps: <i>Q</i> -value: 6.13,						
tcdC117	R-meat	4	16.70	4.15	48.15	- 2.05	0.04	63.48
	Overall	4	16.70	4.15	48.15	- 2.054	0.040	63.48
•	eity between subgrou	ıps: <i>Q</i> -value: 0.00,	p value:1.0					
cdtA	R-meat	2	27.28	12.80	48.95	- 2.05	0.04	0.00
	Seafood	1	30.77	16.20	50.55	- 1.91	0.06	0.00
	Overall	3	29.20	18.12	43.47	- 2.786	0.005	0.00

Table 5 The prevalence of *C. difficile* toxin genes in each sample type

comparing to similar reports [21]. In this study, this difference could be attributed to high consumption of seafood's in diet of Asia and Europe, and a large number of seafood samples have been studied. The lowest prevalence of *C. difficile* was observed in South America.

Most of the studies were on meat and meat products. The contamination of undercooked and prepared foods was evident [24]. The prevalence of *C. difficile* in meat products of this study was the same as a report by Usui 2020 [25], but was lower than study reported from Canada by Warriner in 2017 [26]. It must be noted that the prevalence of food sample isolated C. difficile was so variable with a range of 1.6% from Netherlands [27] to 42% from USA [28]. The prevalence of C. difficile in chicken and poultry meat was 6.2%, which was similar to the previous study (6.7%) [25]. However, the isolation rate of C. difficile from chicken meat samples was ranging from %0 [29-31] to 44.4% in turkey meat samples [32]. It seems that the chicken with skin is more vulnerable to contamination comparing to skin-less chicken samples [33].

Seafood and oysters well-known carriers of *C. difficile* [34]. In the present meta-analysis, the overall contamination rate of seafood was 10.3% and had the highest risk ratio (12.88%). According to another meta-analysis study, pooled prevalence of *C. difficile* in seafood was shown a little bit more in comparison with our pooled prevalence (Seafood risk ratio was 14.3) [21]. This difference may be because of longer time and more included studies. The variation between prevalence of *C. difficile* isolated from seafood's have been seen in many studies from around the world ranged from 3.9% to more than 40% [35–37]. The first report of root vegetables contamination with *C. difficile* was in 1996 [38].

In this study, the overall prevalence of *C. difficile* in contaminated vegetables was 5.7%, which was less than another meta-analysis (12% on average). This would be due to the increase of health level in production and transfer of vegetables [25, 26].

Regardless of the type of food products, the most important issue in relation to *C. difficile* strains is detecting their ribotypes and toxinotypes [24]. Although we could not statistically analyze the *C. difficile* ribotypes data due to vast divergence of the informations, it is obvious that ribotypes 027 and 078 were the most predominants followed by 001 [20, 39], 010 [33] and 020/014 [20] ribotypes. The results of the present study showed that the most common toxinotypes were toxinotypes were toxinotype V, 0, III, respectively. In a review study, the presence of toxin genes in food samples was estimated as 3.5% [32] to 100% [31, 32, 37]. The types of toxinotypes can be important in the development of molecular diagnostic tests and vaccines [40].

As reported by many studies *C. difficile* harboring *tcd*A and *tcd*B, toxin genes were more prevalent than other strains [31].

The contamination risk analysis showed that seafood and RTE-meat are the high-risk foods. While in Rodriguez-Palacios study (21), among different food items, vegetables and seafood were ranked as the high-risk food items, in both studies, seafood is one of the risk food items. This information can be useful for determining preventive food safety measures (cooking food and not consuming raw food) to minimize the possibility of further food contamination.

This study showed that a variety of foods, especially seafood, were at potential risk for *C. difficile*. The frequency of *C. difficile* varied in food samples from different continents. This difference can be attributed to the high consumption of seafood in the diet of Asia and Europe. These results suggest that consumption of raw and undercooked foods is a way to further transmit *C. difficile* to humans.

Conclusions

Therefore, enough cooking of food, suitable washing of animal carcasses in the slaughter process, prevention of carcass contamination with animal feces play an important role in increasing food safety.

Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s41043-023-00369-3.

Additional file 1. The ribotypes of the studies.

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Author contributions

AHA designed the study; SK and SB conducted the data and wrote the manuscript; MR did statistical analyses; RA and JM edited the article; SK, SD, and HM search of articles; all authors read and approved the final manuscript.

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Availability of data and materials

All relevant data are within the manuscript and its Supporting Information files.

Declarations

Ethics approval and consent to participate

The study was approved by ethic committee of Kermanshah University of Medical Sciences (ethic number: IR.KUMS.REC.1398.017). Consent to participate, not applicable.

Competing interests

The authors have declared that no competing interests exist.

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